



## Relative Bioavailability of Lead from Mining Waste Soil in Rats

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The purposes of this study were to determine the extent of absorption of lead (Pb) in mining waste soil from Butte, Montana, and to investigate the effect of mining waste soil dose (g soil/day) on tissue lead concentrations. Young, 7- to 8-week-old male and female Sprague-Dawley rats (5/sex/group) were given mining waste soil that contained 810 or 3908 ppm lead mixed in a purified diet (AIN-76) at four different dose levels (0.2, 0.5, 2, and 5% dietary soil) for 30 consecutive days. Standard groups included untreated controls and dosed feed soluble lead acetate groups (1, 10, 25, 100, and 250  $\mu\text{g Pb/g feed}$ ). The test soil dose levels bracketed a pica child's soil exposure level and the lead acetate concentrations bracketed the test soil dose levels of lead. Liver, blood, and femur were analyzed for total lead concentration using graphite furnace atomic absorption spectroscopy. Clinical signs, body weight, food consumption, and liver weights for test soil and standard groups were similar to control. Tissue lead concentrations from test soil animals were significantly lower than the tissue concentrations for the lead acetate group. Relative percentage bioavailability values, based on lead acetate as the standard, were independent of the two different test soils, dose levels, and sex and were only slightly dependent on the tissue (blood > bone, liver). Mean relative percentage bioavailability values of lead in the Butte mining waste soil were 20% based on the blood data, 9% based on the bone data, and 8% based on the liver data. The results of this study will provide the information needed to determine the significance of lead exposure from Butte soils in assessing human health risks as part of the Superfund Remedial Investigation/Feasibility Study process. © 1992 Society of Toxicology.

This study was initiated to address Superfund health risk assessment issues related to the bioavailability of lead from Butte, Montana, mining waste soils. The sources of lead representing potential risk to human health are the waste rock piles from previous underground mining activities that are scattered throughout the Butte area. Material from these piles

has been used over the years for residential fill material and has, therefore, become indistinguishably mixed with residential soils. An important source of lead exposure for children is the inadvertent ingestion of soil containing lead as a result of normal hand-to-mouth activity and the mouthing of objects that have come in contact with soil containing lead (ATSDR, 1988). Previous research on the bioavailability of lead in ingested soil and dust was summarized by Chaney *et al.* (1989). Chaney (1991) has extended this review and presented the hypothesis that soil in the diet could adsorb lead during passage through the intestinal system. Thus, instead of the generally expected linear effect of dose on concentration of lead in tissue for soluble lead salts added to a purified diet, it is reasonable that the response (tissue lead) should approach a plateau with increasing soil dose. Soil contains hydrous iron oxides, organic matter, and other adsorbing surfaces which may bind lead, thereby reducing absorption in the small intestine. Thus, the inherent chemical properties of soil-lead adsorption sites may reduce the bioavailability of soil-lead compared to soluble lead-salts and lead compounds ingested without soil. This type of pattern was found for plant root uptake of metals from soils with added sewage sludge because the sludge added specific metal adsorption capacity to the soil-sludge mixture (Corey *et al.*, 1987).

The specific objective of this study was to determine the extent of adsorption of lead into the blood, bone, and liver of young male and female Sprague-Dawley rats that were fed various concentrations of lead-contaminated mining waste soil mixed with AIN-76 purified diet for 30 consecutive days. In addition to the mining waste soil treatment groups, a control group (purified diet only) and a standard group (dosed feed lead acetate) were included in the experimental design. At termination, blood, liver, and femur specimens were collected from each animal. Because lead is not homogeneously distributed in the body, but rather is dispersed among several distinct compartments (blood, soft tissue, and bone) (Rabinowitz *et al.*, 1976; Marcus, 1985a,b,c), measuring lead in multiple compartments more accurately reflects total body distribution of lead. Lead concentrations

were analyzed by graphite furnace atomic absorption spectroscopy and/or inductively coupled plasma atomic emission spectroscopy. Relative percentage bioavailability values were estimated by comparing tissue lead concentrations of the test soil to the standard treatment groups. These data provide the information needed to determine the significance of lead exposure from Butte soils in assessing human health risks as part of the Superfund Remedial Investigation/Feasibility Study process.

## METHODS

### Materials

The original test substances were two mining waste soils (Test Soil I, 810 ppm lead; Test Soil II, 8858 ppm lead) that were composites of soils collected from residential areas in Butte, Montana, during November, 1989. Test Soil II had a higher lead concentration than was targeted for this study and, therefore, was blended with Test Soil I to produce Test Soil III which contained 3908 ppm lead. Test Soils I and III were used for the dosed feed preparations. Lead (II) acetate trihydrate ( $(\text{CH}_3\text{CO}_2)_2\text{Pb} \cdot 3\text{H}_2\text{O}$ ) was obtained from Aldrich Chemical Co. (Milwaukee, WI) and was used to prepare the dosed feed formulations for the standard groups.

### Test Substance Composition

**Moisture.** Moisture content was determined by weighing and drying a 5-g sample ( $\pm 0.1$  mg) at  $108^\circ\text{C}$  for 2 hr, followed by reweighing. Percentage moisture was determined by calculating the difference between the predried and dried samples.

**Organic matter.** Percentage organic matter in the test soils was estimated from loss-on-ignition at  $430^\circ\text{C}$  until constant weight or after heating for 24 hr (Davies, 1974). The results after heating for 20 hr are reported because no change in the weight of the sample occurred between 6 and 20 hr.

**pH.** The pH of the test soils was determined using EPA SW 846 Method 9045. The pH was measured with an appropriately calibrated pH meter (Orion, Model 501).

**Total element content.** Total element and lead in the test soils were determined using EPA Test Method SW 846 (Method 3050, Acid Digestion of Sediments, Sludges, and Soils) for sample preparation; and EPA Test Method SW 846 (Method 6010, Inductively Coupled Plasma Atomic Emission Spectroscopy) for metal determination. Method 7000, Atomic Absorption Method, was also used for trace metal analysis by electrothermal furnace technique (EAA).

**Mineralogical evaluation.** Lead mineralogy in Test Soils I and III was determined using a JEOL 8600 electron microprobe (electron beam diameter of  $1\ \mu\text{m}$ ) in the wavelength dispersive mode. Samples were prepared by impregnating the soil with epoxy and polishing the surface with kerosene rather than water to avoid potential dissolution of soluble phases.

**Particle size.** Particle size analysis of the test soils was determined using the electrozone method (Particle Data Laboratories, Ltd., Elmhurst, IL).

### Test System and Animal Maintenance

This was a nonclinical laboratory study performed in compliance with the EPA Good Laboratory Practice Regulations, 40 CFR Part 792 (U.S. EPA, 1989a). Seventy male and 70 female Sprague-Dawley rats (5/sex/dose group; 7–8 weeks of age at initiation of dosing) were supplied by Charles River Laboratories (Kingston, NY). The animals were housed in an environmentally controlled room where the temperature and relative humidity specifications were 19 to  $25^\circ\text{C}$  and 40 to 70%, respectively. Lighting con-

ditions were set to provide 12 hr of fluorescent light (6:00 AM to 6:00 PM) followed by 12 hr of darkness (6:00 PM to 6:00 AM). Animals were individually housed in standard polycarbonate cages, the dimensions of which (width  $\times$  height  $\times$  length) were  $56 \times 32 \times 20$  cm. Cages contained hardwood bedding (Sani-Chips, P. J. Murphy Forest Products, Rochelle Park, NJ). All animals were provided deionized water *ad libitum* ( $<2.0\ \mu\text{g Pb/liter}$ ) by glass bottle reservoirs fitted with stainless steel sipper tubes. All rats were fed *ad libitum* in metal feeders. The untreated control group animals were fed a purified diet (AIN-76 complete meal (Zeigler Brothers, Gardners, PA)). The dosed feed lead treatment group animals were fed AIN-76 sucrose-free meal into which the appropriate amount of sucrose and specific test substance, i.e., test soil or lead acetate, were added. The soil replaced part of the sucrose rather than replacing part of, or diluting, the complete mixed diet. The AIN-76 complete meal and AIN-76 sucrose-free meal contained  $<0.20\ \mu\text{g Pb/g}$ .

### Dosing Regimen and Administration

Dosed feed formulations were prepared for the animals fed lead acetate and the test soils mixed in the diet. The dose levels selected for use on this study were based on reported soil exposure levels for pica children. For a 15-kg child with pica-for-soil, approximately 10 g of soil and 250 g dry diet are estimated to be consumed daily (U.S. EPA, 1989b). Based on these values, soil constitutes approximately 4% of the pica child's diet. Thus, dose levels of 2 and 5% soil in the diet were selected in this study to bracket the pica child exposure level. The lower dose levels of 0.2 and 0.5% soil reflected logarithmic decreases from the higher dose levels to the lowest exposure level believed to produce tissue concentrations that are above the analytical detection level. Typical children (exclusive of those with pica-for-soil) ingest less than 100 mg of soil per day, on average, which is equivalent to approximately 0.04% soil in their diet (Calabrese *et al.*, 1989). However, feeding rats a comparable dose would result in tissue lead concentrations indistinguishable from background concentrations. For the Test Soil I group, the 0.2, 0.5, 2, and 5% soil doses corresponded to 1.62, 4.05, 16.2, and 40.5 ppm lead. For the Test Soil III group, the concentration of lead in the four soil dose levels was 7.82, 19.5, 78.2, and 195 ppm. The exposure levels of the lead acetate groups (1, 10, 25, 100, and 250 ppm) were chosen to bracket the estimated exposure levels for the two dosed feed test soil groups. Dosed feed concentrations, stability, and homogeneity were verified during the study. All doses were homogenous with relative standard deviations of less than 5%, except the 1-ppm lead acetate preparation. Actual concentrations ranged from 93 to 116% of the target concentrations. Appropriate dosed feed mixtures were presented to the rats at approximately the same time each day.

### In-Life Parameters

Animals were observed twice daily for any signs of moribundity or mortality and once daily for signs of toxicity. Individual body weight determinations were made once weekly and individual food consumption determinations were made daily.

### Tissue Collection and Preparation

Animals were euthanized with a single overdose injection of sodium pentobarbital. Blood was collected into a syringe by a cardiac puncture. Animals were necropsied and liver and bone (right femur) were collected, weighed, and frozen at approximately  $-20^\circ\text{C}$ . Cardiac blood specimens were transferred to heparinized containers and refrigerated until the time of preparation for analysis. Each blood sample was well mixed on a rotating tumbler for at least 30 min prior to removing an aliquot of known weight for lead analysis. Liver specimens were homogenized with a tissue homogenizer (Brinkman Polytron, Westbury, NY) and frozen (approximately  $-20^\circ\text{C}$ ) until analysis. The whole bone was placed in 2 N NaOH to digest any soft residual tissue, dried, and weighed. The defleshed bone was then placed in 6 N HCl until it

was completely solubilized. The completely solubilized bone specimens were frozen (approximately  $-20^{\circ}\text{C}$ ) until analysis.

#### *Determination of Lead in Biological Fluids, Tissues, Soils, and Dosed Feed Mixtures*

Samples were analyzed using a Perkin-Elmer Model 5000 Zeeman atomic absorption spectrophotometer equipped with a Model 500 graphite furnace, (GFAAS) or an inductively coupled plasma atomic emission spectrometer (ICP-AES) (Thermo Jarrell-Ash Model 61-975) for low or high concentrations of lead, respectively. Blood specimens were diluted with 0.05% Triton X-100 in 0.2% (v/v)  $\text{HNO}_3$ . Aliquots (20  $\mu\text{l}$ ) of the test solutions were injected onto the platform of the graphite furnace with 5  $\mu\text{l}$  of matrix modifier solution (0.4% monobasic ammonium phosphate and 0.14% magnesium nitrate hexahydrate in 1% (v/v)  $\text{HNO}_3$ ). Solubilized bone specimens were digested in 15 M  $\text{HNO}_3$  in a boiling water bath, diluted with water, and injected into the graphite furnace with a matrix modifier solution (same as blood except also containing 1% calcium). Liver samples were digested with three parts 15 M  $\text{HNO}_3$  to one part 70%  $\text{HClO}_4$  and lead was determined by graphite furnace as described previously for blood. Actual soils and dosed feed mixtures were digested with 15 M  $\text{HNO}_3$ , 30%  $\text{H}_2\text{O}_2$ , and 12 M  $\text{HCl}$  on a hot plate. The digested samples were redissolved in 15 M  $\text{HNO}_3$  and diluted to a proper concentration within the linear range of the instrument.

Lead in all samples was calculated from linear regression equations using the method of standard additions (Klein and Hach, 1977). The method detection limits (MDLs) for the blood, bone, and liver lead concentrations were 11.1  $\mu\text{g/liter}$ , 0.56  $\mu\text{g/g}$ , and 0.04  $\mu\text{g/g}$ , respectively. The method detection limit is defined as the lead concentration that yields an absorbance equal to three times the standard deviation of a sample with a concentration of lead that is distinctly detectable above, but close to, the blank absorbance measurement. All lead concentrations below the MDL were set at the detection limits for statistical analysis.

#### *Statistics*

The principal objectives of the statistical analysis were to characterize the nature of the trends in tissue lead concentrations with increasing delivered concentrations of lead in the diet and to characterize the variability of the responses about these trends in tissue uptake. Regression models were fitted to the data to quantify the dose-response trends and to provide smoothed estimates of the tissue uptake concentrations and their variability.

Nonlinear regression models were fitted to the results from the lead acetate and test soil groups to describe the relations between the delivered dose (mg Pb/kg BW) and tissue lead concentrations ( $\mu\text{g/liter}$  or mg/kg). Each model simulates the dose-response relationship, plateauing as the input dose increases. Separate fits were carried out for males and for females. This resulted in six fits per tissue type.

The regression models related the group mean tissue lead concentrations to the external-delivered lead dose. The data suggested that the tissue concentrations increased to a limiting value (i.e., an asymptote) as the external lead dose increased and maintained a nonzero level even in the absence of lead in the diet (i.e., a background level).

For blood lead concentrations, the model fitted the data ( $p > 0.05$ ) in five of the six cases; there was borderline lack of fit ( $p = 0.04$ ) for the lead acetate males. For bone lead concentrations, the model fitted each treatment and sex combination. For liver lead concentrations, the model fitted the lead acetate groups for both males and females. There was insufficient lead uptake in the test soil groups to warrant fitting regression models. Tissue concentrations were estimated based on the unsmoothed average concentrations among the five animals per sex at each dose.

Each animal had background levels of lead in its blood, bone, and liver apart from any lead that was fed to it during the study. These background levels were adjusted for when the amounts of dosed lead taken up from the study diet were estimated. For the liver lead concentrations, the low dose

concentrations for each treatment and sex were, for the most part, at or below the detection limit, 0.04  $\mu\text{g/g}$ . The background level for liver was, therefore, set at the detection limit, 0.04  $\mu\text{g/g}$ , for both males and females.

The relative bioavailability for a particular dose of a test soil was defined as the ratio of the lead uptake from that dose of the test soil to the lead uptake of the same dose of the lead acetate treatment. The lead uptake estimates from the test soil and from the lead acetate standard were adjusted for background levels before being compared. The regression curves for the lead acetate groups were interpolated to obtain predictions at doses corresponding to the test soil group doses.

## RESULTS

### *Test Soil Characterization*

Test soil characterization results (lead concentration, percentage moisture content, percentage organic matter, pH, total element content, mineralogic evaluation, and particle size) for Test Soils I and III are summarized in Table 1. Total lead concentrations in the two test soils mixed in the feed and fed to the animals were  $810 \pm 21$  ppm for Test Soil I and  $3908 \pm 31$  ppm for Test Soil III which are typical of many mining waste soils. The percentage organic matter content for Test Soils I and III was 3.0 and 4.1, respectively. The pH of the soils was strongly to extremely acidic (according to USDA-SCS Soil pH Categories), with pH values of approximately 4.5 and 3.7 for Test Soils I and III, respectively, which are typical of noncalcareous soils derived from mining waste. Several elements were present at high concentrations that may have affected lead absorption. These included, aluminum, arsenic, cadmium, calcium, copper, iron, manganese, and zinc.

The lead phase mineralogy provides a useful indicator of the important mineral and noncrystalline phases controlling lead solubility of ingested soils. In Test Soil I, 45% of the lead mass was present as anglesite ( $\text{PbSO}_4$ ) and galena ( $\text{PbS}$ ). Coronadite ( $\text{MnPbMn}_2\text{O}_4$ ) and lead oxide phosphate ( $\text{Pb}_4\text{O}(\text{PO}_4)_2$ ) accounted for 50% while iron elyite ( $(\text{Cu-Fe})\text{Pb}_4(\text{SO}_4)\text{OH}_4$ ) accounted for the remaining 5%. Galena is a common ore mineral while anglesite is an oxidation product that forms a rind around galena in acid environments produced by dissolution of pyrite in mining waste rock (Nordstrom, 1982) (Fig. 1). The low lead concentration in Test Soil I resulted in a limited number of lead-bearing crystals in the sample. Test Soil III, however, contained a large number of lead minerals, resulting in better counting statistics and a more representative distribution. The lead mineralogy observed in Test Soil III consisted primarily of anglesite and galena (77%) with a variety of minor phases. Both samples displayed alteration products and rinding (coating or armoring of the original mineral grain by a reaction product) of the primary lead phases, e.g., galena ( $\text{PbS}$ ) altering to anglesite ( $\text{PbSO}_4$ ) (Fig. 1), cerussite ( $\text{PbCO}_3$ ) altering to anglesite, lead oxide ( $\text{PbO}_2$ ) altering to anglesite, and further alteration of anglesite to plumbojarosite

TABLE 1  
Test Soil Characterization

Analysis	Test soil I	Test soil III
Lead concentration (ppm) <sup>a</sup>	810 ± 21	3,908 ± 31
% Moisture <sup>b</sup>	1.7 ± 0.1	1.8 ± 0.2
% Organic matter <sup>c</sup>	3.0 ± 0.1	4.1 ± 0.2
pH <sup>d</sup>	4.50 ± 0.01	3.67 ± 0.02
Particle size (GMS ± GSD) <sup>e</sup>	48 ± 46 μm	42 ± 44 μm
Composition (ppm) <sup>f</sup>		
Aluminum	23,500 ± 600	17,300 ± 200
Arsenic	65 ± 2	1,380 ± 20
Cadmium	12.3 ± 0.3	23.0 ± 1.0
Calcium	5,270 ± 955	6,940 ± 40
Copper	490 ± 60	1,490 ± 30
Iron	45,700 ± 1650	69,300 ± 450
Manganese	3,450 ± 70	2,200 ± 100
Zinc	5,750 ± 140	6,040 ± 95
Mineralogic analysis <sup>g</sup>		
Anglesite (PbSO <sub>4</sub> )	28%	53%
Galena (PbS)	17%	24%
Coronadite (MnPbMn <sub>2</sub> O <sub>4</sub> )	28%	—
Lead oxide phosphate (Pb <sub>2</sub> O(PO <sub>4</sub> ) <sub>2</sub> )	22%	—
Iron elyite ((Cu-Fe)Pb <sub>2</sub> (SO <sub>4</sub> ) <sub>2</sub> (OH) <sub>2</sub> )	5%	—
Barite ((Pb-Ba)SO <sub>4</sub> )	—	5%
Lead phosphate (Pb <sub>2</sub> (PO <sub>4</sub> ) <sub>3</sub> )	—	4%
Thorium-lead phosphate (Th-Pb(PO <sub>4</sub> ) <sub>3</sub> )	—	4%
Plumboferrite (PbFe <sub>2</sub> O <sub>7</sub> )	—	4%
Plumbojarosite (PbFe <sub>4</sub> (SO <sub>4</sub> ) <sub>2</sub> (OH) <sub>12</sub> )	—	3%
Lead oxide	—	3%

<sup>a</sup> The concentration of lead in the soils was determined by ICP-AES using EPA SW 846 Method 6010. Triplicate aliquots of the soils were digested according to EPA Method 3050 and duplicate aliquots of each digestate were analyzed. Tabled values are reported as means ± standard deviation.

<sup>b</sup> Percentage moisture content was determined by differences in soil weight before and after drying. Tabled values reflect the mean ± standard deviation of duplicate replicates.

<sup>c</sup> Organic matter content was estimated from loss-on-ignition at 430°C after heating for 20 hr (Davies, 1974). Tabled values are reported as the mean ± standard deviation of duplicate analyses.

<sup>d</sup> The pH of the soil was determined using EPA SW 846 Method 9045. Tabled values are reported as the mean ± standard deviation of duplicate analyses.

<sup>e</sup> Particle size analysis of the test soils was determined using basic electrozone technology.

<sup>f</sup> Total element content was determined by ICP-AES using EPA SW 846 Method 6010. EPA Method 3050 was used for digestion of samples. Values (ppm) are the mean of triplicate samples ± standard deviation.

<sup>g</sup> Mineral identification was conducted using a JEOL 8600 electron microprobe. Values reflect results of a single analysis and are expressed as a percentage of the total lead content.

(PbFe<sub>2</sub>(SO<sub>4</sub>)<sub>2</sub>(OH)<sub>12</sub>). In addition, galena encapsulated in a silicate or pyrite matrix was often observed. Precipitation of K-jarosite (KFe<sub>3</sub>(SO<sub>4</sub>)<sub>2</sub>(OH)<sub>6</sub>) from the dissolution of pyrite which surrounds and armors anglesite was pervasive in both soils. The nonlead heavy mineral fraction consisted of barite (BaSO<sub>4</sub>), pyrite (FeS<sub>2</sub>), sphalerite (ZnS), and iron oxide

(Fe<sub>2</sub>O<sub>3</sub>). Due to the predominance of PbSO<sub>4</sub> as the lead-bearing phase in Test Soil III and the rinding of minor Pb phases by PbSO<sub>4</sub> alteration products, PbSO<sub>4</sub> is likely to control lead solubility in both test soils under gastrointestinal tract conditions. It is important to note that dissolution of PbSO<sub>4</sub> will also be kinetically hindered by the rind of jarosite that is stable in acidic media (pH < 4, Vlek *et al.*, 1974) and that will further limit lead mineral exposure to GI tract fluids.

The geometric mean size (GMS) and geometric standard deviation (GSD) were determined using volume-based diameter distribution data. The GMS (±GSD) values were 48 μm (±46) for Test Soil I and 42 μm (±44) for Test Soil III. These results indicated that the particle size of the test soils were similar to the size of ingested soil particles commonly found to adhere to children's hands (<100 μm) (Duggan *et al.*, 1985; Chaney *et al.*, 1989).

### Signs of Toxicity

Treatment group animals were similar in appearance and behavior to the control group animals and no overt signs of toxicity were observed.

### Body Weight Determinations

There were no statistically significant ( $p < 0.05$ ) dose-dependent or treatment-dependent changes in body weight or body weight gain when compared to control group values for either sex. Body weight gain results indicated that animals were thriving and growing during exposure to lead from either the lead acetate or soil lead mixed in the diet. After 4 weeks, group mean body weight gain values ranged from 27 to 40 g/week for the males and 6 to 18 g/week for the females. Thus, exposure occurred during a rapid growth phase and the lead acetate and soil lead did not compromise growth patterns.

### Food Consumption Determinations

The overall group mean food consumption values for the untreated control group rats during the 30-day in-life period was 23 g feed/day/animal. Group mean food consumption values for all treatment groups were not significantly different ( $p < 0.05$ ) from the control group values for both sexes. Although there were a few treatment groups with subgroups that had elevated group mean food consumption values, the increases were generally slight in magnitude and did not reflect dose-dependent changes. In addition, the data suggested that no palatability problems occurred with the lead acetate or soil lead dosed feed formulations.

### Daily Exposure Index

The group mean daily exposure index values (milligrams lead consumed per kilogram body weight per day) by study

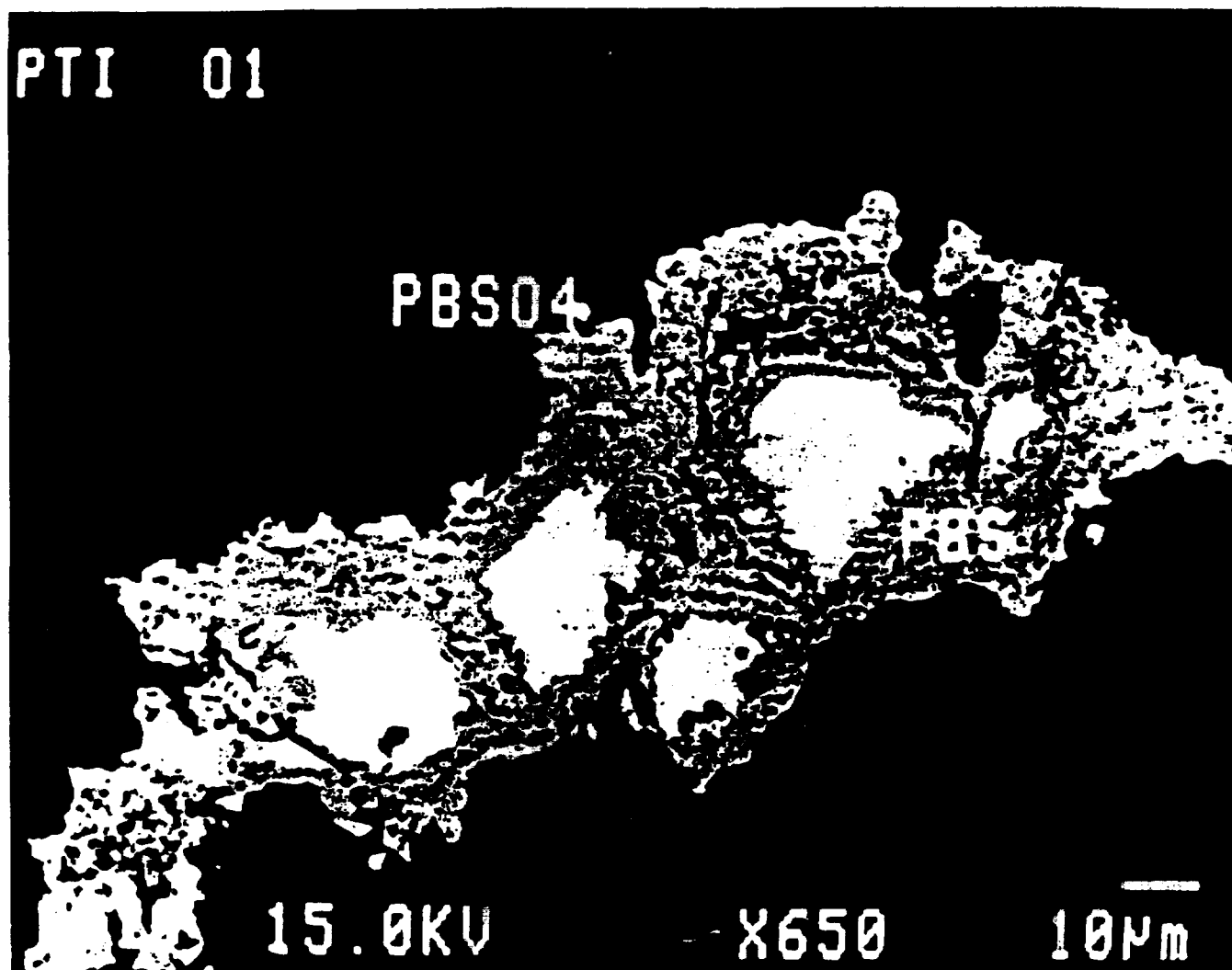


FIG. 1. Galena ( $\text{PbS}$ ) oxidizing to a rind of anglesite ( $\text{PbSO}_4$ ).

week for male and female rats are summarized in Table 2. The group mean daily exposure index data indicated approximately proportional increases in the level of exposure (absolute amount and per body weight) at the different dose levels for each treatment group and both sexes. The exposure index differed slightly between the sexes for many of the dose levels of each treatment group, with the exposure level being higher for females than for males. The sex difference was attributed to similar group mean daily food consumption values between the sexes but substantially lower body weight values for the female animals when compared to the male animals.

#### *Tissue Lead Levels*

**Blood.** Overall, the group mean whole blood lead concentration values for the Test Soil I and III groups were sig-

nificantly lower than the blood lead concentration values for comparable exposure levels of the lead acetate group (Fig. 2). The group mean whole blood lead concentration values increased with increasing dose levels for all treatment groups but were not proportional, reaching a plateau at the higher dose levels in the soil treatment groups. A plateau was less apparent for the male and female lead acetate groups. For the most part, similar group mean blood lead concentration values were observed for male and female rats within the same treatment group.

**Bone.** The group mean bone lead concentration values increased with increasing dose levels for all treatment groups (Fig. 3). Bone lead levels were very low following dietary ingestion of Test Soils I and III compared to bone lead levels after ingestion of lead acetate. Lead absorption and distri-

TABLE 2  
Daily Exposure Index Summary<sup>a</sup>

Treatment group	Targeted dose level (ppm)	Daily exposure index (mg Pb/kg BW)									
		Males					Females				
		Week 1	Week 2	Week 3	Week 4	$\bar{x} \pm SD$	Week 1	Week 2	Week 3	Week 4	$\bar{x} \pm SD$
Lead acetate	1	0.0890	0.0820	0.0730	0.0590	$0.0758 \pm 0.0129$	0.107	0.0990	0.0920	0.0760	$0.0935 \pm 0.0102$
	10	0.851	0.712	0.757	0.653	$0.743 \pm 0.084$	0.926	0.806	0.908	0.790	$0.858 \pm 0.059$
	25	1.79	1.63	1.86	1.73	$1.75 \pm 0.10$	1.87	1.73	2.14	1.94	$1.92 \pm 0.17$
	100	7.30	7.02	7.48	7.06	$7.22 \pm 0.22$	10.1	9.05	10.9	8.84	$9.72 \pm 0.96$
	250	20.3	17.3	14.0	13.4	$16.3 \pm 3.2$	26.6	29.0	22.9	24.2	$25.7 \pm 2.7$
Test soil I	1.62	0.150	0.132	0.102	0.0930	$0.119 \pm 0.026$	0.163	0.167	0.116	0.135	$0.145 \pm 0.024$
	4.05	0.365	0.333	0.273	0.236	$0.302 \pm 0.058$	0.528	0.560	0.466	0.450	$0.501 \pm 0.052$
	16.2	1.20	1.10	1.14	1.08	$1.13 \pm 0.05$	1.89	1.91	1.84	1.65	$1.82 \pm 0.12$
	40.5	3.70	3.60	2.93	2.70	$3.23 \pm 0.49$	4.90	4.34	4.42	3.51	$4.29 \pm 0.58$
Test soil III	7.82	0.618	0.615	0.728	0.605	$0.642 \pm 0.058$	0.909	0.822	0.895	0.896	$0.881 \pm 0.040$
	19.5	1.84	1.66	1.44	1.43	$1.59 \pm 0.20$	2.18	2.27	2.24	2.28	$2.24 \pm 0.05$
	78.2	5.33	5.03	5.18	4.98	$5.13 \pm 0.16$	6.79	7.07	7.56	8.14	$7.39 \pm 0.59$
	195	12.8	12.1	11.7	11.7	$12.1 \pm 0.5$	24.8	22.2	23.1	22.7	$23.2 \pm 1.1$

<sup>a</sup> Dose levels were normalized to mg of lead (Pb)/kg of body weight (BW) for the dosed feed groups. Normalization was performed using the following equation: Daily exposure index (mg Pb/kg BW) = (group mean daily lead consumption (mg), week<sub>4</sub>/group mean body weight (kg), week<sub>4</sub>). Daily lead consumption values were calculated by multiplying the concentration of lead in the dosed feed times the daily food consumption.

bution into the bone was detected after administration of lead acetate at all dose levels tested but only at the higher Test Soil I and III dose levels tested. The increases in bone lead concentrations were not proportional to increasing dose levels. The bone lead concentrations appeared to plateau at the higher dose levels for all treatment groups, although the plateau was much less apparent for the male lead acetate group. Similar group mean bone lead concentrations were observed for male and female rats within the same treatment group, except for the lead acetate treatment group, whereby the male group mean bone lead concentrations were approximately threefold higher than the females.

**Liver.** The group mean liver lead concentration values increased with increasing dose levels for the lead acetate and Test Soil III groups (Fig. 4). Overall, the mean liver lead concentration values for the Test Soil I and III groups were lower than the liver lead concentration values for the lead acetate groups. The increase in liver lead concentrations was not proportional to increasing dose levels but appeared to plateau at the higher dose levels for the lead acetate and Test Soil III groups. Again, the plateau for the lead acetate male group was much less apparent than the other groups. Several of the animals in the 1-ppm lead acetate group; the 0.2, 0.5, and 2.0% Test Soil I groups; and the 0.2 and 0.5% Test Soil III groups had mean liver lead concentrations that were slightly above or not different from background levels. Similar group mean liver lead concentration values were observed for male and female rats within the same treatment group,

except for the lead acetate treatment group, whereby the male group mean liver lead concentrations were approximately twofold higher than the females.

#### Relative Percentage Bioavailability

**Blood.** Relative percentage bioavailability at the low dose levels (0.2 and 0.5%) could not be accurately determined due to the very low blood lead concentrations attained at these exposure levels. Only at the higher dose levels, i.e., 2 and 5%, were blood lead levels following lead acetate and test soil ingestion elevated sufficiently above background for precise estimation of bioavailability values (Table 3). The 2 and 5% dietary soil dose levels bracket the pica child's daily soil ingestion. For Test Soils I and III, relative percentage bioavailability values ranged from 12 to 26% and 20 to 27%, respectively, for the male and female 2 and 5% dose level groups based on the lead acetate standard. There were no statistically significant differences between the dose levels or sexes. Based on blood data, the overall mean relative percentage bioavailability value for Test Soils I and III at 2 and 5% soil in the diet was approximately 20%.

**Bone.** Similar to the blood, relative percentage bioavailability values for the bone data at the low dose levels (0.2 and 0.5% dietary soil) were highly variable. Data for the 2 and 5% dose levels provided more reliable relative percentage bioavailability results (Table 3). For Test Soils I and III, relative percentage bioavailability values ranged from approximately 5 to 11% and 8 to 13%, respectively, for the male

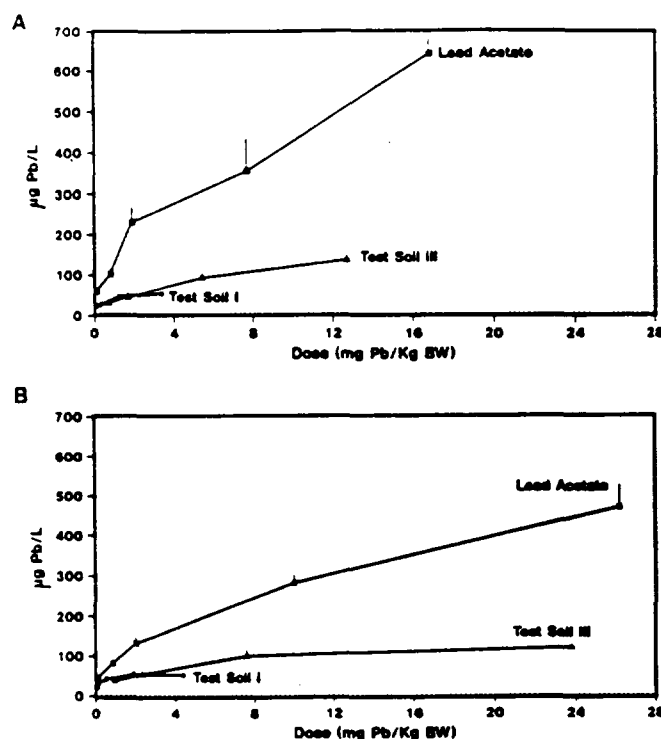


FIG. 2. Lead concentration in blood ( $\mu\text{g Pb/liter}$ ) versus dose ( $\text{mg Pb/kg BW}$ ) in (A) male and (B) female rats. Data are expressed as the mean  $\pm$  standard deviation,  $n = 5$  (duplicate analyses per animal). Method detection limit is  $11.1 \mu\text{g Pb/liter}$ .

and female 2 and 5% dose level groups based on the lead acetate standard. There were no statistically significant differences between dose levels or sexes. Using bone data, the overall mean relative percentage bioavailability value for Test Soils I and III at 2 and 5% soil in the diet was approximately 9%.

**Liver.** For Test Soil I, liver lead levels were slightly above or at method detection limits at dose levels of 0.2, 0.5, and 2% dietary soil for both sexes. Thus, the extent of absorption and eventual accumulation of lead in the liver could not be accurately determined. This resulted in relative percentage bioavailability estimates for these dose levels that were highly variable and imprecise. At the 5% dose level, the relative percentage bioavailability values were approximately 9% (males) and 8% (females) based on the lead acetate standard (Table 3). For Test Soil III, liver lead levels were near method detection limits at dose levels of 0.2 and 0.5% dietary soil for both sexes. Thus, relative percentage bioavailability estimates for the 0.2 and 0.5% dose levels were variable due to relatively low absorption and accumulation of lead to the liver following ingestion of Test Soil III. For the male 2 and 5% Test Soil III dose levels, group mean relative percentage bioavailability values were approximately 7 and 8%, respec-

tively, and, for the females, 14 and 10%, respectively, based on the lead acetate standard (Table 3). The overall mean relative percentage bioavailability value for Test Soils I and III at 2 and 5% soil in the diet was approximately 8%.

## DISCUSSION

This study was conducted to determine the extent of absorption (relative percentage bioavailability) of lead from two different mining soils using young Sprague-Dawley rats fed soil mixed with a purified diet. It is the first study to fully investigate the bioavailability of lead in soils containing mine waste using a soil dose-response approach. Male and female Sprague-Dawley rats (5 animals/sex/group) were fed mining waste soil (810 and 3908 ppm lead) from Butte, Montana, mixed in an AIN-76 purified diet at four dose levels for 30 consecutive days. The soil was mixed with a purified diet to lower the background levels of lead found in control animals and to allow the detection of the soil lead in the animal's tissues even at low lead levels. This diet also maximizes lead absorption because it is relatively low in calcium. Low dietary calcium increases lead absorption because calcium and lead are absorbed through similar mechanisms (Six and Goyer,

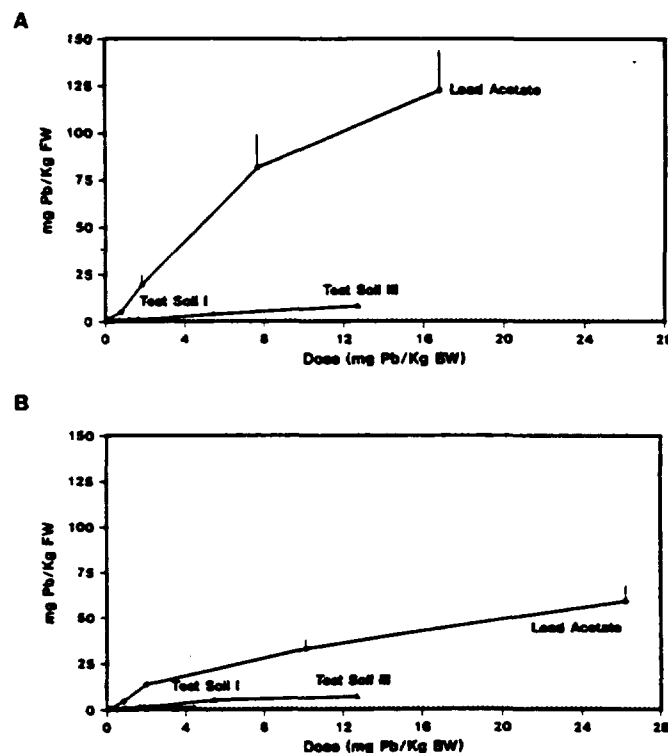


FIG. 3. Lead concentration in bone ( $\text{mg Pb/kg FW}$ ) versus dose ( $\text{mg Pb/kg BW}$ ) in (A) male and (B) female rats. Data are expressed as the mean  $\pm$  standard deviation,  $n = 5$  (duplicate analyses per animal). Method detection limit is  $0.56 \text{ mg Pb/kg}$ .

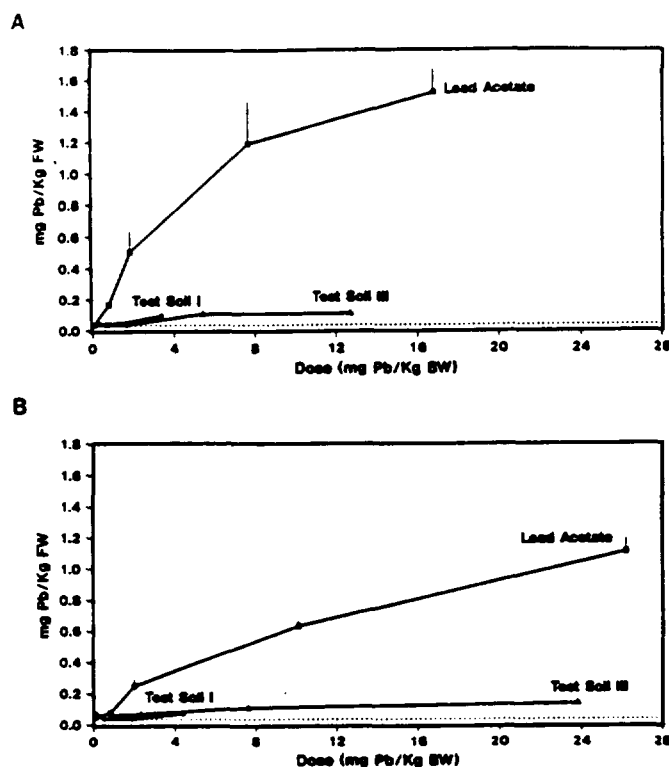


FIG. 4. Lead concentration in liver (mg Pb/kg FW) versus dose (mg Pb/kg BW) in (A) male and (B) female rats. Data are expressed as the mean  $\pm$  standard deviation,  $n = 5$  (duplicate analyses per animal). Method detection limit is 0.04 mg Pb/kg.

1970; Mahaffey, *et al.*, 1973; ATSDR, 1988). Regular rat chow attenuates the availability of lead so strongly that other properties of the soils which might affect lead absorption by humans from ordinary diets are masked by the strong lead binding or precipitative action of the chow diet. The nutritionally complete purified AIN-76 diet (Bieri *et al.*, 1977) has much less ability to inhibit absorption. In addition, purified diets more closely simulate the low fiber diets of most U.S. citizens than do rat chow diets.

The rat was used because it is the preferred species for metabolism and pharmacokinetic studies performed under the Toxic Substances Control Act (TSCA) regulations (U.S. EPA, 1982) and because it is considered an acceptable model for human risk assessment (National Research Council, 1983). In addition, the food consumption patterns and stomach pH of rats and young children are similar (Chaney, 1991). For example, rats and young children generally eat intermittently and frequently while they are awake, and the pH in the rat stomach ranges from 2.6–5.1 which is similar to the pH range measured in children's stomachs after they have eaten. The age range and developmental state of the rats in this study also support their use in prediction of lead uptake in children. The rats used were 7–8 weeks old at the

start of dosing and exhibited rapid growth as indicated by their increase in body weight. The males gained about 125 g (36% increase) and the females about 45 g (21% increase) over the duration of the study. At the start of dosing, these animals were not sexually mature because vaginal patency generally occurs at about 5 to 6 weeks of age while sperm production is observed at about 9 to 10 weeks. In addition, pregnancy cannot be sustained until an age of at least 10 to 12 weeks (personal communication, Joanne Killinger, 1991). Rats of this age also absorb calcium very well with as much as 30–49% of a single dose of calcium being deposited in the bone after oral administration of calcium salts in tablet form (Kenny, 1981; Killinger *et al.*, 1986).

The study was 30 days in length because of the long biological half life of lead. Thirty days appeared to be an adequate compromise between the need for sufficient time for accumulation of lead in the blood and tissue, while still balancing the need for an exposure period which ensured that animals remained in a rapid growth phase. The target tissues measured were whole blood, liver, and bone, representing

TABLE 3  
Relative Percentage Bioavailability Values of Lead for Male and Female Rats Administered Test Soils I and III Mixed with Feed<sup>a</sup>

Group <sup>b</sup>	Test soil I	Test soil III
Blood		
Males		
2%	18.1 (6.0)	19.6 (3.3)
5%	12.1 (3.6)	21.5 (3.8)
Females		
2%	25.7 (7.8)	26.8 (4.8)
5%	13.8 (4.7)	22.1 (4.4)
Bone		
Males		
2%	8.0 (3.5)	7.5 (1.4)
5%	4.8 (1.5)	7.5 (1.4)
Females		
2%	10.6 (3.3)	13.3 (2.2)
5%	6.1 (1.9)	13.0 (2.6)
Liver		
Males		
2%	4.3 (2.4)	7.1 (1.5)
5%	8.7 (2.0)	7.5 (1.5)
Females		
2%	0.6 (3.1)	13.6 (3.1)
5%	8.2 (2.8)	9.8 (2.1)

<sup>a</sup> Relative percentage bioavailability values were determined using the lead acetate group as the standard. Tabled data are reported as the mean with the standard error of the mean in parentheses.

<sup>b</sup> 2 and 5% refer to the percentage soil in the diet.

the important compartments for lead distribution. In addition, the chronic feeding design allowed us to keep dietary lead levels similar to those of pica children, and to keep the ratio of lead to other elements close to those of these children. Other study designs have used 10 times or higher dietary lead for shorter periods, potentially confounding interactions between lead and other elements such as calcium and phosphorous.

The clinical appearance, body weight, food consumption, and liver weight values in the soil-treated animals were similar to those in the control group animals, indicating no overt toxicity that may have affected the relative percentage bioavailability values estimated for this study. Significantly higher lead concentrations were measured in the blood, bone, and liver of animals fed lead acetate when compared to animals fed the test soils mixed in diet. Thus, under comparable dosing conditions, the bioavailability of lead in the Butte soils was considerably less than that of lead acetate administered in the diet. Over the dose range tested (0.2 to 5% soil in diet), the blood, bone, and liver lead concentration versus dose profiles generally plateaued for animals of both sexes when fed the test soils mixed in diet. The plateau response to soil lead is extremely apparent if one compares the pattern of response of tissue lead for the soil animals to the high linear slope at the start of the lead acetate curves. These data are consistent with results from epidemiological studies that show there is a weak relationship between soil lead concentration and human blood lead levels at other mining sites with a high proportion of lead sulfide in the mineral assemblage (Bornschein *et al.*, 1990).

Relative percentage bioavailability values, based on lead acetate as the standard, were independent of the two different test soils, dose levels, and sex, and only slightly dependent on the tissue (blood > bone, liver). It is appropriate to compare lead acetate with the forms of lead in mine waste soils because lead acetate is highly soluble (thus maximizing bioavailability) and can serve as a surrogate for soluble lead compounds ingested in the human diet. Overall relative percentage bioavailability values (assuming no biologically significant differences between sexes, dose levels, or soils) were 20% based on the blood data; 9% based on the bone data; and 8% based on the liver data (2 and 5% soil dose levels only). The low bioavailability of lead in the Butte soil-treated animals agreed favorably with low blood lead levels (average of 3.5  $\mu\text{g}/\text{dl}$ ) found in children from Butte, Montana (Butte-Silver Bow County Environmental Health Study, 1991). Negligible lead absorption (i.e., only slightly above background concentrations) occurred at test soil dose levels of 0.2 and 0.5%. These lower dose levels, although more closely approximating soil lead exposure to that of normal children, were still above the average lead level intake of a child. Thus, uptake of lead from soil at dose levels similar to those of a

normal child's exposure would not be detectable under this study protocol.

This study demonstrates that lead in the Butte mining waste soil is less bioavailable than lead from an automotive or paint lead source. For example, rats fed soil lead from a roadside (auto exhaust lead) and from along the edge of a lead-paint house demonstrated lead concentrations in blood, bone, liver, kidney, and brain after 30 days that were similar to rats fed lead acetate; while at 90 days, tissue concentrations in bone and kidney were about 70 to 80% those in lead acetate rats (Dacre and Ter Haar, 1977). Chaney *et al.* (1984) reported relative percentage lead bioavailability values (based on bone) in Baltimore garden soils ranging from 15 to 70% (average of 33.4%) of the dietary lead acetate. Based on bone lead, the bioavailability of lead in Butte soils was about one-quarter that in the Baltimore garden soils. These comparisons underscore the importance of evaluating lead source and species when predicting bioavailability values for lead in soils. Soil mineralogy is a critical parameter in assessing lead dissolution because the different lead-bearing solids characteristic of different sources have varying solubilities under the different pH regimes of the stomach prior to and after ingestion of food. The low bioavailability is due in part to the pervasive encapsulation and alteration of the common lead-bearing solids (galena and anglesite) that preclude ready dissolution of these minerals over their residence time in the GI tract. Mineralogy of the Butte soils demonstrates that galena has oxidized to anglesite, with peripheral precipitation of K-jarosite coating the lead grains. This armoring process serves to inhibit dissolution of lead-bearing solids both physically through a reduction in the surface area of anglesite exposed to GI tract fluids and chemically due to the less soluble nature of K-jarosite in an acidic medium, i.e.,  $\text{pH} < 4$  (Vlek *et al.*, 1974; Davis *et al.*, 1992).

The bioavailability of lead is influenced by the species of lead incorporated into the soil (which varies depending on the source of lead), the size of the lead-containing soil particles, the matrix incorporating the lead species, and nutrients or other compounds ingested with the lead (Steele *et al.*, 1990; Chaney *et al.*, 1989). The toxicokinetics of lead can be affected by interactions with essential elements. Increased levels of iron decrease lead uptake (Barton *et al.*, 1978). Low dietary levels of zinc (Cerklewski and Forbes, 1976; El-Gazzar *et al.*, 1978) and copper (Klauder *et al.*, 1973; Klauder and Petering, 1975) increase lead absorption. High levels, therefore, may act to impede the absorption of lead (Edshall and Wyman, 1958; Underwood, 1977; Brewer *et al.*, 1985). Thus, high levels of aluminum, arsenic, cadmium, calcium, copper, iron, manganese, and zinc may have affected lead absorption, indicating the importance of determining the elemental content and mineralogy of the soils.

The lead in the original Butte soils may actually be even less bioavailable than the results of the present experiment

show. The test soils were passed through a 250- $\mu\text{m}$  sieve and blended to obtain a more homogeneous lead concentration. However, the sieved portion of the soil (<250  $\mu\text{m}$  fraction) represented only about 10% of the original soil sample, with the remaining 90% of larger particle size. In addition, by mechanically mixing the soil into the feed, the soil particle sizes were reduced further. For example, Test Soil I had a geometric mean volume-based diameter of 48  $\mu\text{m}$  ranging from 1 to 194  $\mu\text{m}$  while Test Soil III had a geometric mean volume-based diameter of 42  $\mu\text{m}$  ranging from 1 to 182  $\mu\text{m}$ . Thus, the actual mean particle size of the test soils was well below the 250- $\mu\text{m}$  sieve size. In general, the lower the particle size, the greater the absorption of lead because smaller particles (higher surface area to mass) will dissolve more rapidly in the GI tract, thus producing more solubilized lead. Decreasing the particle size of the test soil, therefore, facilitates the absorption of lead into the systemic circulation. Thus, the bioavailability of lead in the indigenous Butte soils is likely to be less than that measured in the present study due to larger particle sizes found in the original Butte soil samples. In this study, the mean particle size of both test soils was below the upper limit for the size of soil particles reported to adhere to children's hands (<100  $\mu\text{m}$ ) that constitutes the major route of ingestion (Duggan *et al.*, 1985; Chaney *et al.*, 1989). Therefore, the bioavailability of lead in the indigenous Butte soils may be less than that determined for the present study for a pica child because the particle sizes of the test soils in this study were of a size which mimicked reported soil particle size values for normal human exposure but also optimized the dissolution rate and extent of absorption. Thus, for a child with pica-for-soil, this study represents a worst case scenario because pica children ingest large particles during bulk soil ingestion and not exclusively the < 100  $\mu\text{m}$  size fraction.

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## REFERENCES

- Agency for Toxic Substances and Disease Registry (ATSDR) (1988). *Toxicological Profile for Lead*, draft. ATSDR, Public Health Service, U.S. Department of Health and Human Services, Atlanta, GA.
- Barton, J. C., Conrad, M. E., Nuby, S., and Harrison, L. (1978). Effects of iron on the absorption and retention of lead. *J. Lab. Clin. Med.* 92, 536-547.
- Bieri, J. G., Stoewsand, G. S., Briggs, G. M., Phillips, R. W., Woodard, J. C., and Knapka, J. J. (1977). Report of the American Institute of Nutrition ad hoc Committee on Standards for Nutrition Studies. *J. Nutr.* 107, 1340-1348.
- Bornschein, R., Clark, S., Pan, W., and Succop, P. (1990). Midvale Community Lead Study Final Report. Department of Environmental Health, University of Cincinnati Medical Center, Cincinnati, OH.
- Brewer, G. J., Hill, G. M., Dick, R. D., Prasad, A. S., and Cossack, Z. T. (1985). Interactions of trace elements: Clinical significance. *J. Am. Coll. Nutr.* 4, 33-38.
- Butte-Silver Bow County Environmental Health Study (1991). Conducted by the Butte-Silver Bow County Health Department and the University of Cincinnati, Department of Environmental Health.
- Calabrese, E. J., Barnes, R., Stanek, E. J., III, Pastides, H., Gilbert, C. E., Veneman, P., Wang, X., Laszity, A., and Kostecki, P. T. (1989). How much soil do young children ingest: An epidemiologic study. *Regul. Toxicol. Pharmacol.* 10, 123-137.
- Cerklewski, F. L., and Forbes, R. M. (1976). Influence of dietary zinc on lead toxicity in the rat. *J. Nutr.* 106, 689-696.
- Chaney, R. L. (1991). Soil lead chemistry in relation to bioavailability of lead in soil and dust. In *Proc. Symp. Bioavailability and Dietary Uptake of Lead* (R. Cothorn and R. W. Elias, Eds.), Research Triangle Park, NC.
- Chaney, R. L., Mielke, H. W., and Sterrett, S. B. (1989). Speciation, mobility, and bioavailability of soil lead. *Environ. Geochem. Health* 11(Suppl.) 105-129.
- Chaney, R. L., Sterrett, S. B., and Mielke, H. W. (1984). The potential for heavy metal exposure from urban gardens and soils. In *Proceedings, Symposium on Heavy Metals in Urban Gardens* (J. R. Preer, Ed.), pp. 37-84. Agriculture Experimental Station, University of the District of Columbia, Washington, DC.
- Corey, R. B., King, L. D., Lue-Hing, C., Fanning, D. S., Street, J. J., and Walker, J. M. (1987). Effects of sludge properties on accumulation of trace elements by crops. In *Land Application of Sludge—Food Chain Implications* (A. L. Page, T. J. Logan, and J. A. Ryan, Eds.), pp. 25-51. Lewis Publishers, Michigan.
- Dacre, J. C., and Ter Haar, (1977). Lead levels in tissues from rats fed soils containing lead. *Arch. Environ. Contam. Toxicol.* 6, 111-120.
- Davies, B. E. (1974). Loss-on-ignition as an estimate of soil organic matter. *Soil Sci. Soc. Am. Proc.* 38, 150.
- Davis, A., Ruby, M. V., and Bergstrom, P. D. (1992). Bioavailability of arsenic and lead in soils from the Butte, Montana, mining district. *Environ. Sci. Technol.* 26(3), 461-468.
- Duggan, M. J., Inskip, M. J., Rundle, S. A., and Moorcroft, J. S. (1985). Lead in playground dust and on the hands of school children. *Sci. Total Environ.* 44, 65-79.
- Edshall, J. T., and Wyman, J. (1958). *Biophysical Chemistry*, Vol. 1, p. 591. Academic Press, New York.
- El-Gazzar, R. M., Finelli, V. N., Boiano, J., and Petering, H. G. (1978). Influence of dietary zinc on the lead toxicity in rats. *Toxicol. Lett.* 1, 227-234.
- Kenny, A. D. (1981). *Intestinal Calcium Absorption and its Regulation*. CRC Press, Boca Raton, FL.
- Killinger, J. M., Pignone, N. D., Chin, T. Y., and Freudenthal, R. I. (1986). The disposition and relative bioavailability of calcium from various calcium salts in the rat. *Pharmacol. Res.* 3 (Supplement), 165S.
- Klauder, D. S., Murthy, L., and Petering, H. G. (1973). Effect of dietary intake of lead acetate on copper metabolism in male rats. In *Trace Substances in Environmental Health. VI. Proceedings of University of Missouri's 6th Annual Conference on Trace Substances in Environmental Health* (D. D. Hemphill, Ed.), pp. 131-136. Univ. Missouri Press, Columbia, MO.
- Klauder, D. S., and Petering, H. B. (1975). Protective value of dietary copper and iron against some toxic effects of lead in rats. *Environ. Health Perspect.* 12, 77-80.

- Klein, R., and Hach, C. (1977). Standard additions, uses and limitations in spectrophotometric analysis. *Am. Lab.* 9, 21-30.
- Mahaffey, K. R., Goyer, R., and Haseman, J. K. (1973). Dose-response to lead ingestion in rats fed low dietary calcium. *J. Lab. Clin. Med.* 82, 92-100.
- Marcus, A. H. (1985a). Multicompartment kinetic models for lead. I. Bone diffusion models for long-term retention. *Environ. Res.* 36, 442-458.
- Marcus, A. H. (1985b). Multicompartment kinetic models for lead. II. Linear kinetics and variable absorption in humans without excessive lead exposure. *Environ. Res.* 36, 459-472.
- Marcus, A. H. (1985c). Multicompartment kinetic models for lead. III. Lead in blood plasma and erythrocytes. *Environ. Res.* 36, 473-489.
- National Research Council (1983). *Risk Assessment in the Federal Government: Managing the Process*. Committee on the Institutional Means for Assessment of Risks to Public Health, Commission on Life Sciences, National Academy Press, Washington, D.C.
- Nordstrom, D. K. (1982). Aqueous pyrite oxidation and the consequent formation of secondary iron minerals. In *Acid Sulfate Weathering* (J. A. Kittrick, D. S. Fanning, and L. R. Hossner, Eds.), pp. 37-56. Soil Science Society of America, Madison WI.
- Rabinowitz, M. B., Wetherill, G. W., and Kopple, J. D. (1976). Kinetic analysis of lead metabolism in healthy humans. *J. Clin. Invest.* 58, 260-270.
- Six, K. M., and Goyer, R. A. (1970). Experimental enhancement of lead toxicity by low dietary calcium. *J. Lab. Clin. Med.* 76, 933-942.
- Steele, M. J., Beck, B. D., Murphy, B. L., and Strauss, H. S. (1990). Assessing the contribution from lead in mining waste to blood lead. *Regul. Toxicol. Pharmacol.* 11, 158-191.
- Underwood, E. J. (1977). In *Trace Elements in Human and Animal Nutrition*, 4th ed., Vol. 70, pp. 133, 205. Academic Press, London.
- United States Environmental Protection Agency (1982). *Health Effects Test Guidelines and Support Documents*. Report No. EPA 560/6-82/001.
- United States Environmental Protection Agency (1986). Office of Solid Waste and Emergency Response. SW 846. Test Methods for Evaluating Solid Waste.
- United States Environmental Protection Agency (1989a). Federal Register: Toxic Substances Control Act (TSCA). Good Laboratory Practice Standards: Final Rule. 40 CFR Part 792.
- United States Environmental Protection Agency (1989b). *Development of Risk Assessment Methodology for Land Application and Distribution and Marketing of Municipal Sludge*. Report No. EPA 600/6-89/001.
- Vlek, P. L. G., Blom, Th. J. M., Beek, J., and Lindsay, W. L. (1974). Determination of the solubility product of various iron hydroxides and jarosite by the chelation method. *Soil Sci. Soc. Am. Proc.* 38, 429-432.